## Gene expression databases

Sean Eddy, PhD

# Outline

- How to measure gene expression?
   Microarrays/RNA-seq
- What are gene expression databases?
- Which ones exist?
- How do they differ?
- How can they be used?
- What needs to be taken care of?
- What can I do with specialized databases?

# Microarrays: the beginning of high throughput gene expression



Nature Reviews | Drug Discovery

## http://www.nature.com/nrd/journal/v1/n12/ images/nrd961-f1.gif

- Compartmentalized chips with sequences bound to the surface
- Sample is applied to surface
- Hybridizes to complementary sequence
- Hybridizations are quantified
  - No binding, "no" signal
  - Can only find what is specifically searched for
  - Usually represented as n x m matrix

# RNA-seq: the future of high throughput gene expression



Weihua Zeng & Ali Mortazavi

Nature Immunology 13, 802–807 (2012) | doi:10.1038/ni.2407 Published online 21 August 2012

- Samples (cDNA libraries) are applied to a sequencer
- Millions of sequences are generated and mapped onto a genome
- Sequence reads are quantified
  - No sequence = no expression
  - Can find novel transcripts, splice isoforms, fusion genes, etc.
- Quality of mapping depends largely on library prep and decisions made on how RNA is initially processed.

# Gene expression databases

- Repositories for gene expression data
  - Mostly microarray and now RNAseq
  - Primarily for storage
  - Curated or un-curated
  - Access to data on different levels:
    - Datasets
    - Individual levels
- Integrated databases
  - Contain array data and additional data of the samples
  - Array data tends to be more annotated
  - More analytical tools
  - Smaller (more QC and curation needed)
  - Often no direct data access

# Why do they exist

- Transparency/reproducibility of publications
  - Journals require data to be available for analysis
  - Nowadays raw data is required
  - Databases offer single resource and standardized access
- Data was generated for a specific purpose, but is not limited to that purpose
  - Can be reanalyzed in a different context
  - Can be combined with other datasets
  - Can be used as independent validation

# Gene expression repository examples

- Gene expression omnibus (www.ncbi.nlm.nih/geo/)
  - <u>1,117,462</u> samples, <u>3848</u> datasets

S NCB	Resources 🗹 How To 🗹							Sigr	<u>in to NCBI</u>
GEO H	ome Documentation 🔻	Query & Browse 🔻	Email GEO						
GEO is a sequence	public functional genomics	Omnibus data repository supportin iools are provided to help	g MIAME-compl users query and	iant data submi d download exp	issions. Array- and eriments and curated	gene		Gene Expressio	on Omnibus
expressio	n profiles.						Keyword or GEO A	Accession	Search
						_			

Array express (www.ebi.ac.uk/arrayexpress/)

ArrayExpress – functional genomics data

ArrayExpress Archive of Functional Genomics Data stores data from high-throughput functional genomics experiments, and provides these data for reuse to the research community.

Browse ArrayExpress

Jata Content

Updated today at 12:00

- 64933 experiments
- 1968713 assays
- 40.97 TB of archived data
- Princeton University MicroArray database (PUMAdb)

- 40084 experiments, 6598 made public

• NCBI SRA, ENA and Princeton HTseq for NGS data

# What is in a gene expression database?

- Gene expression data in different forms:
  - Resolution:
    - Gene level
    - Transcript level
    - Exon level
      - And / or raw data
  - Comprehensiveness
    - Targeted arrays
    - Whole genome arrays
  - Different platforms (microarrays, RNAseq)
- Generally only gene expression, may have limited sample information

# Where does the data come from?

- Expression profiles of
  - Patients
  - Model systems
  - Cell cultures
- Data used for publication
  - Most journals now require raw data submission
  - Very coarse quality control (peer review)
  - QC depends mostly on authors
- Datasets submitted without publication
   Little or no QC
- Most datasets are tailored towards a specific question

# Example: GEO GSE32591

- Go to <a href="http://www.ncbi.nlm.nih.gov/geo/">http://www.ncbi.nlm.nih.gov/geo/</a>
- Enter GSE32591 into search box
- Click on "Analyze with GEO2R"
  - How would you set up the groups for analysis?
  - What do you get?
    - Does that make sense? How can results be verified?
- Go to "value distribution" tab
  - What do you see?
  - What are possible explanations?



👜 🛫 🗴 👧 TMN:

# What can be done with GEO?

# What can be done with GEO?

- Programmatic access for data download
  - <u>http://www.ncbi.nlm.nih.gov/geo/info/geo\_paccess.html</u> (GEO)
  - <u>http://www.ebi.ac.uk/arrayexpress/help/</u> programmatic\_access.html (ArrayExpress)
- Pre-computed analyses and on the fly analyses
  - Search by gene across all GEO experiments
  - Search by experiment to retrieve cluster analysis
  - Search by gene sequence for matching expression profiles
    - Described by Barret and Edgar, Methods Mol. Biol. 2006 "Mining Microarray Data at NCBI's Gene Expression Omnibus (GEO)"
      - <u>http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1619899/</u>



# What questions can be answered?

# What questions can be answered?

- If you download: anything
  - Only limited by your knowledge, skills, resources
- Pre-computed results
  - Preselected analysis methods/ sample groups
  - Generally within one dataset
- On-the-fly analyses
  - Sets of genes that cluster in under conditions given
  - Sample properties may not be entirely transparent.

# What can be answered by doing it yourself?

# What can be answered by doing it yourself?

- The quality of the data
  - Is part of the data low quality?
  - Does some of the data not fit into the set (e.g. batch effect, outliers for other reasons)
  - Is it adequately processed?
- What is the relationship between expression data and non-expression variables?
  - How does my gene (of interest) associated with experimental treatments, clinical parameters?
- What are patterns across datasets?
  - Does my finding hold up across similar analyses in independent datasets?

# Why do you have to do it yourself?

- Quality control:
  - QC parameters are often glossed over in papers and in micraorray submissions
  - For Affymetrix QC modules are available, freely available and widely accepted in the bioinformatic community
  - Other array types have distinct, but also similar properties
  - http://www.nature.com/nbt/focus/maqc/index.html
- Relations to non-expression data variables
  - Data is often not standardized within fields

# Why not?

- Analysis across datasets:
  - Because:.... How?
  - Need to find a common standard for identification
  - Values need to be made comparable
    - If absolute expression values used, dynamic range can be a problem
    - Is ratios used, information about expression level lost
  - Non-expression data even worse

# Who is the target group for doing it yourself?

- Users with experience in expression data
  - Crucial information (STUFF) is missing



# Why is this a problem?

 Excludes investigators with good hypotheses but lacking bioinformatic skills



"Must be a clinical fellow."

"The computer says I need to upgrade my brain to be compatible with microarray data analysis."

# How to fix that?



# How to fix that?

- Specialized databases
  - Datasets are easier to find
    - Datasets relevant to specific areas are collected in one place
      - NephroSeq for renal disease
      - Oncomine for cancer
  - Datasets are standardized and expertly curated
    - Controlled vocabulary is introduced for non-expression data
    - Curation of expression possible by introducing standardized references and data transformations across datasets
      - Gene IDs/Gene Symbols as references
      - Z-transformation or median centering of log transformed expression data

# Nephroseq (<u>www.nephroseq.org</u>)

## Nephroseq

.ogin

login	
USER ID:	
PASSWORD:	
Forgot password?	[
Not a user? Register now!	L

### about

Nephroseq is supported by the Applied Systems Biology Core as part of the University of Michigan O'Brien Renal Center. The primary goal of the Applied Systems Biology Core is to provide to the renal research community a platform for integrative data mining of comprehensive renal disease gene expression data sets, in order to:

 Define molecular characteristics/features in the circulation or kidney and associate them with known disease phenotypes so as to obtain a better understanding of the pathophysiology of a specific renal disease

2. Identify markers of disease progression and treatment response (i.e., biomarkers)



### Welcome to Nephroseq

Developed for the renal research community, Nephroseq is a platform for integrative data mining of genotype/phenotype data, with optimized workflows that lead from search to visualization and from question to answer to next question:

The expression of a gene is highly correlated with well-known podocyte genes. Is the gene functionally important in glomeruli?

L Contact

- A gene is significantly differentially expressed in a subset of disease patients. Is the gene associated with a certain phenotype, severity or sub-category of the disease?
- A set of genes is significantly up-regulated in disease patients. Are the disease genes inversely related to the target profile of a compound/drug?

#### ABOUT NEPHROSEQ

Originally a collaborative effort, Nephroseq is now solely developed and maintained by the Applied Systems Biology Core at the University of Michigan. This resource combines a wealth of publicly available renal gene expression profiles - gathered and curated by an experienced team of data scientists, bioinformaticians, and nephrologists - with a sophisticated analysis engine and powerful web application designed for data mining and visualization of gene expression data.

Nephroseq provides researchers with a rich set of publicly available renal gene expression data, packaged with the tools and interface necessary to analyze it, all aimed at seeking answers to questions and advancing a molecular understanding of kidney disease to ultimately improve clinical outcomes.

In particular, Nephroseq provides unique access to datasets from the Personalized Molecular Nephrology Research Laboratory incorporating clinical data which is often difficult to collect from public sources.

# Oncomine (www.oncomine.com

## www.oncomine.org)

		Qupgrade @ contact
login	Oncomine <sup>™</sup> Research Edition: 715 datasets and 86,733 sample	les
USER ID: PASSWORD: • Forgot password? • Not a user? Register now! Inews Visit our newsroom to find out about the latest developments at Thermo Fisher	Design better experiments. Gain more insights. Prepare to publish faster.	
Scientific. Did you know that you can include the fee for Oncomine Research Premium Edition in US federal grant applications? Contact us for more information.	With Oncomine <sup>™</sup> Research Premium Edition, you can: Design better experimentsAnswer more questions with fewer experiments, select the most promising gene or cell line, and test your hypothesis.	iontorrent by Thermo Fisher Scientific
events	Gain more biological insightsDiscover novel targets for therapeutic development, interrogate gene expression profiles, and identify drug and biological interactions. Prepare to publish fasterValidate your results faster, visualize data easier and make	The Origin of the Oncomine Platform The Oncomine Platform was conceived by physicians, scientists, and software engineers at the University of Michigan. It was commercialized by Aruf Chinnaiyan and Dan Bhordes in February 2006 with the pool of building a
Whether it's a conference, trade show, or webinar, you will find us at these events.	The Oncomine <sup>®</sup> Platform-from web applications to translational bioinformatics services -provides solutions for individual researchers and multinational companies, with robust, peer-reviewed analysis methods and a powerful set of analysis functions that compute gene expression signatures, dusters and gene-set modules, automatically extracting biological insights from the data. It has become an industry-standard tool cited in more than 1,100 peer-reviewed journal articles. The Oncomine Platform has been used as a foundation for ground-breaking discoveries with unique features that include:	version that would have a greater ability to impact drug development and clinical practice.
	<ul> <li>Scalability – with 700+ independent datasets</li> <li>High quality – with expertly curated data</li> </ul>	
	Consistency – with a rich, extensive and controlled ontology of terms	

 Standardized analysis – with conventions that assure clear and consistent interpretations of results

Oncomine Research Edition remains free to the academic and nonprofit cancer research communities.

## NephroSeq and Oncomine

- Pros:
  - Each focus on one area of interest
  - Clinical data for many individual samples available
  - Advanced analysis using integrated systems biology tools in a pre-defined automated manner
  - Meta analysis possible
  - User friendly, free accessible for academic users
  - Hypotheses-generating
- Cons:
  - No raw data download
  - No programmatic access
  - Only predefined analyzes



# **Two Search Options**

• Gene specific search:

– Gene

- Dataset search:
  - Specific conditions/diseases

## NPHS2: encodes podocin, a podocyte specific protein

# Gene Search

## Gene summary view



# Gene Search

## Gene summary view



22 out of 33 analysis meet your threshold for NPHS2 in 2 out of 2 datasets

# Four Basic Analysis Modes

- Differential expression
- Co-expression analysis
- Outlier analysis

Heterogeneity within predefined groups

• Concepts analysis

– Gene set (Nephromine & third-party sources)

# Gene Search

## Differential expression (Box graph)

NEPHROMINE		Welcome, Wenjun Ju. 🛠 tools 😤 help 🗱 logout	
search	datasets Concepts ORDER BY: Under-expression: Gene Rank	visualize Differential Analysis OTHER VIE	IEWS: 🕨
filter	ON:         NPHS2           THRESHOLD BY:         P-VALUE           P-VALUE         FOLD CHANGE           0.05         1.5	GROUP BY: TISSUE Type SHOW: Only Samples in Analysis  + PRIMARY CONCEPT: EXP NPHS2 Expression in Lindenmeyer Normal Tissue	Port: 🕨
Gene: NPH52     Analysis Type: Tissue Type Analysis     Dataset Type: Normal Tissue Panel	Compare   Clear All	Panel Tissue Type: Glomeruli vs. Tubulointerstitium Lindenmeyer Normal Tissue Panel Statistics	
Primary Filters T - Analysis Type Coexpression Analysis (2) - Differential Analysis (2)	Tissue Type: Glomeruli vs. Tubulointerstitium a=5.68E-8 fold change = -10.623 694 Roth Normal Tissue Panel (353)	Under-expression Gene Rank: 694 (in top 6%) P-value: 5,68E-8 Reporter: 220424_at t-Test: -26.283 Fold Change: -10.623	
+ Demographics Analysis (1) Tissue Type Analysis (2) Outlier Analysis (2) + Group	Issue Type: Urerna vs. Namey           p = 1.03E-4         fold change = -12.091         865           Tissue Type: Bronchus vs. Kidney           p = 1.57E-4         fold change = -11.538         881           Tissue Type: Lymph Node vs. Kidney		
+ Tissue Type + Dataset Type Sample Filters ↓ Dataset Filters ↓	p = 1.84E-4 fold change = -10.851 937 Tissue Type: Papilla of the Tongue vs. Kidney p = 3.23E-4 fold change = -8.761 954 Tissue Type: Ovary vs. Kidney p = 2.27E-4 fold change = -9.745 964		
Concept Filters 🛓	Tissue Type: Prostate Gland vs. Kidney           p = 3.38E-4         fold change = -9.433         965           Tissue Type: Thyroid Gland vs. Kidney           p = 2.46E-4         fold change = -9.546         970	in the second	
	Tissue Type: Trachea vs. Kidney     p = 2.66E-4 fold change = -9.255 996 ■     Tissue Type: Breast vs. Kidney     p = 1.70E-4 fold change = -10.594 1151 ■     Tissue Type: Topoli vs. Kidney	0.5	
	p = 2.23E-4         Fold change = -9.932         1172           Tissue Type: Adrenal Cortex vs. Kidney           p = 2.87E-4         fold change = -9.031         1174           Tissue Type: Lung vs. Kidney	1. Glomeruli (6) 2. Tubulointerstitium (6) Lindenmeyer Normal Tissue Panel	
	p = 2.05E-4 fold change = -10.044 1281 🔲 💌	PLoS One 2010/07/12         12 samples         NPHS2 Information           mRNA         12,624 measured genes         Reporter Information           Human Genome U133A Array         Human Genome U133A Array         Human Genome U133A Array	

# THE HUMAN PROTEIN ATLAS

ABOUT & HELP

SEARCH ? »	
NPHS2	Search Clear Fields »
e.g. CD44, ELF3, KLK3, or use Fields to search specific fields such as protein_class:Transcription factors or chromosome:X	



# Gene Search

## Correlation with clinical continuous variation



## Gene Search Outlier analysis

Outlier analysis helps to identify an expression profile where differential pattern is only seen in <u>a fraction of samples</u> of all patients within a disease type.

<u>Why do we need it</u>: 25% of patients show over-expression of a gene. This gene may not generate a significant p-value in a t-test comparing DN relative to normal kidney.

<u>How to do it</u>: Transform all samples within a dataset, so that genes could be ranked by their expression from high to low. The data transformation is performed at certain percentile bins (75, 90 & 95%), and a line is drawn at the percentile of that analysis to define outliers.

For example, in an outlier analysis at the 75th percentile, the system draws a line at the point at which only the top 25th percentile samples extend above it.

## Gene Search Outlier analysis



## Differential expression – Dataset search

search	4	da	tasets	concepts
	Г	ORD	ER BY: Dataset Name	
			ON:	
filter			▶ Compare   Clear All	
selected 16 datasets (1121 samples)	Ι.		Flechner Transpla	ant (62)
Analysis Type: Differential Analysis			Cadaveric Donor Kidney Rejection vs. No Rejection	Specimen: Acute
	1	<b>-</b>	Cadaveric Donor Kidney	Specimen: Age
Primary Filters 🕴 🕇			Cadaveric Donor Kidney	Specimen: GFR (MDRD)
<ul> <li>Analysis Type</li> </ul>			Cadaveric Donor Kidney	Specimen: Sex
Coexpression Analysis (16)			Cadaveric Donor Periphe Specimen: Acute Rejection	ral Blood Lymphocyte on vs. No Rejection
Differential Analysis (16)     Demographics Analysis (14)			Cadaveric Donor Periphe Specimen: Age	eral Blood Lymphocyte
Donor Type Analysis (1)			Cadaveric Donor Periphe Specimen: GFR (MDRD)	eral Blood Lymphocyte
Group Analysis (9) + Indices Analysis (3)			Cadaveric Donor Periphe Specimen: Renal Dysfund	eral Blood Lymphocyte
+ Pathology Analysis (11)			Cadaveric Donor Periphe	ral Blood Lymphocyte
Tissue Type Analysis (8) + Treatment Analysis (1)		Ē	Cadaveric Donor Tissue 1	Type: Kidney vs.
Outlier Analysis (16)		÷	Kidney Specimen Donor T	uper Living up
+ Group		1	Cadaveric	ype, civing vs.
+ Donor Type		É.	Living Donor Kidney Spec	imen: Age
+ Tissue Type		Ē.	Living Donor Kidney Spec	imen: GFR (MDRD)
+ Dataset Type			Living Donor Kidney Spec vs. No Rejection	imen: Renal Dysfunctio
Sample Filters 🛓			Living Donor Kidney Spec	imen: Sex
Dataset Filters ↓ Concept Filters ↓			Living Donor Peripheral E Specimen: Age	Blood Lymphocyte
			Living Donor Peripheral B Specimen: GFR (MDRD)	Blood Lymphocyte
		Ē	Living Donor Peripheral B Specimen: Renal Dysfunc	Blood Lymphocyte tion vs. No Rejection
			Living Donor Peripheral B Specimen: Sex	Blood Lymphocyte

- cyte ocyte cyte tion cyte unction
- tion
- Living Donor Tissue Type: Kidney vs. Peripheral

#### visualize **Differential Analysis**

### GROUP BY: Group (Cadaveric Donor Kidney Specimen) SHOW: Only Samples in Analysis

#### 1 | 2 | 3 | 4 | 5 »

Rank

٠

#### Comparison of All Genes in Flechner Transplant Over-expression in Cadaveric Donor Kidney Specimen: Acute Rejection vs. No Rejection

(log2 median-centered intensity)

ank	P-value	Fold Change	Gene								Reporter	Gene
1	2.26E-8	1.51	NIPAL3								37850_at	NIPAL3
2	2.20E-7	1.41	ST×BP5L								34130_at	STXBP5L
з	3.33E-7	1.51	KSR1								1716_at	KSR1
4	3.53E-7	1.51	SRC								1938_at	SRC
5	4.04E-7	1.68	LLGL1								804_s_at	LLGL1
6	4.45E-7	1.73	FSTL4								34518_at	FSTL4
7	6.21E-7	1.43	ARID 3A								35913_at	ARIDBA
8	7.71E-7	2.02	CYP1A1								1024_at	CYP1A1
9	7.93E-7	1.44	RIN1								1777_at	RIN1
10	9.63E-7	1.63	PDGFB								1573_at	PDGFB
11	1.10E-6	2.14	SKI								1918_at	SKI
12	1.28E-6	1.52	COL11A2								1027_at	COL11A2
13	1.36E-6	1.81	PTCH1								836_at	PTCH1
14	1.93E-6	1.63	ZNF646								39863_at	ZNF646
15	2.13E-6	1.55	COL2A1								598_at	COL2A1
16	2.22E-6	1.42	KISS1								1645_at	KISS1
17	2.29E-6	1.39	TBL3								41603_at	TBL3
18	2.31E-6	1.49	CYP2C18								1477_s_at	CYP2C18
19	2.44E-6	1.47	MAST1								35962_at	MAST1
20	2.52E-6	1.34	PCD HGC3								657_at	PCDHGC3
21	3.14E-6	1.40	BRF1								141_s_at	BRF1
					1			2	,			

OTHER VIEWS: >

EXPORT:

Over-expression

ZDC

+ PRIMARY CONCE

#### Legend

**T** 

- 1. No Rejection (5)
- 2. Acute Rejection (6)



Note: Colors are z-score normalized to depict relative values within rows. They cannot be used to compare values between rows.

#### Elechner Transplant

Am J Transplant 2004/09/01	62 samples
mRNA	8,603 measured genes
Human Genome 11954-Av2 Array	

# Differential expression – dataset search – compare analysis

- Compare different analyzes
- Data is standardized on upload (centered to 0 and standardized by variance)

+ PRIMA

Gene

NIPAL3

STX8P5L

KSR1

SRC

LLGL1

FSTL4

ARIDBA

CYP1A1

RIN1

SKL

PDGFB

PTCH1

ZNE646

COL2A1

CYP2C18

PCDHGC3

MAST1

BRF1

KISS1

TBL3

COL11A2

Reporter

37850 at

34130\_at

1716 at

1938\_at

804\_s\_at

34518\_at

35913\_at

1024\_at

1777 at

1573\_at

1918 at

1027\_at

836 at

598 at

1645 at

41603\_at

1477\_s\_at

35962\_at

141\_s\_at

657\_at

39863 at

all features are mapped to common identifier (EntrezGeneID)

search		datasets	concepts		visual	ize							
	L E L	ORDER BY: Dataset Name	2		Differe	ntial Ana	lysis						
<u>्</u>			-		GROUP BY:	Group (Cada	veric Donor Kidney	Specimen)				-	
					SHOW-	Only Samples	in Analysis 💌						
filter		🄄 Compare   Clear A	LL CONTRACTOR OF										
selected 16 datasets (1121 samples)		Flechner Trans	plant (62)	-	1   2   3	3   4   5 ×				_			
X Analysis Type: Differential Analysis		Cadaveric Donor Kidn Rejection vs. No Reje	ey Specimen: Acute ction		Over-	Compa expression in	Cadaveric Donor Ki	enes in Fle idney Specimer	echner n: Acute	Reject	ion vs. N	<b>t</b> io Rejec'	tior
		Cadaveric Donor Kidn	ey Specimen: Age				(logz media	in-centered in	censicy)				
Primary Filters		Cadaveric Donor Kidn	ey Specimen: GFR (MDRD)		Rank	P-value	Fold Change	Gene	_		_	_	
- Analysis Type		Cadaveric Donor Kidn	ey Specimen: Sex		1	2.26E-8	1.51	N IPAL3					
Coexpression Analysis (16)		Cadaveric Donor Peri	pheral Blood Lymphocyte		2	2.20E-7	1.41	ST×BP5L				4	_
<ul> <li>Differential Analysis (16)</li> </ul>		Specimen: Acute Reje	ction vs. No Rejection		з	3.33E-7	1.51	KSR1					_
+ Demographics Applyris (14)		Cadaveric Donor Peri Specimen: Age	pheral Blood Lymphocyte		4	3.53E-7	1.51	SRC					_
Denor Tupo Applysis (14)		Cadaveric Donor Peri	pheral Blood Lymphocyte		5	4.04E-7	1.68	LLGL1					_
Converting (R)		Specimen: GFR (MDRD	)		6	4.45E-7	1.73	FSTL4					-
Group Analysis (9)		Cadaveric Donor Peri	pheral Blood Lymphocyte		7	6.21E-7	1.43	ARID 3A					-
+ Indices Analysis (3)		Specimen: Renal Dystu	Inction vs. No Rejection		8	7.71E-7	2.02	CYP1A1					
Tierre Tree Analysis (11)		Specimen: Sex	pheral Blood Lymphocyte		9	7.93E-7	1.44	RIN1					-
Tissue Type Analysis (8)		Cadaveric Donor Tiss	ue Type: Kidney vs.		10	9.63E-7	1.63	PDGFB					-
+ Treatment Analysis (1)	l I e	Peripheral Blood Lymp	hocyte		11	1.10E-6	2.14	SKI					-
Outlier Analysis (16)		Kidney Specimen Dong	or Type: Living vs.		12	1.28E-6	1.52	COLITAZ					-
+ Group		Cadaveric			13	1.305-0	1.61	ZNEEAE					
+ Donor Type		Living Donor Klaney S	pecimen: Age		14	2.135.6	1.05	2017040					
+ Tissue Type	LL C	Living Donor Kidney S	pecimen: GFR (MDRD)		15	2.136-0	1.55	KISS1					
+ Dataset Type	1 L L	vs. No Rejection	pecimen: Renal Dysfunction		17	2.22E 0	1.72	TRUB					
Sample Filters 🚽		Living Donor Kidney S	pecimen: Sex		18	2.25E 0	1.55	CYP2C18					
Dataset Filters 🛓		Living Donor Peripher	al Blood Lymphocyte		19	2 44F-6	1 47	MAST1					
Concept Filters ↓		Specimen: Age			20	2.52E-6	1.34	PCDHGC3					
		Living Donor Peripher Specimen: GFR (MDRD	al Blood Lymphocyte		21	3.14E-6	1.40	BRF1					
		Living Donor Peripher Specimen: Renal Dysfu	al Blood Lymphocyte Inction vs. No Rejection							1			2
		Living Donor Peripher	al Blood Lymphocyte		1	egend	(5)						
		Living Dopor Ticcup Ti	uper Kidney vs. Periphorol	-	2	. Acute Reier	ction (6)						

# Meta analysis

- Find out which genes are significantly more expressed in glomeruli compared to tubulointerstitium
- Can you verify that with another dataset?
- Or with more than one other dataset?
- Does it matter if the datasets are different?
- Can you imagine a use of this functionality for an exclusive filter (NOT)

# Example



# **Concepts Analysis**

**Concepts** are sets of genes representing some aspect of biology.

Concepts are derived from both **Nephromine gene expression signatures** as well as **third-party sources** such as Gene Ontology, KEGG Pathways, Human Protein Reference Database, etc.

User can upload a self-defined custom concept (a set of genes) to Nephromine to explore it's association with Nephromine and third-party concepts.

# **Concepts Analysis**

	-		_	
NEPHROMINE		v	Unload Custom	
-				reoncept
Nephromine	e Overview		Manage My Co	
NEW IN NEP	HROMINE :: MARCH 2012		Change passwo	ord
Rephromine: Upload My Concept - Windows Internet	et Explorer			nalyses are specified below.
http://www.nephromine.org/resource/ui/tool/concept.html	ACTION=SHOW_UPLOAD_CONCEPT		<b>•</b>	
Upload My Concept				e glomerulus. Genes with 'a priori' N1, DAG1, DDN, EHD3, MYH9, NES,
	Podo-50-symbol			
Concept Name:		Download	list from C-tools	erular and tubulointerstitial
Gene Set (Text File):		to the deal	ton thon unlos	strong enrichment for
Category: HUGO Ger	ne Symbol	to the desi	ctop, then uploa	iu
Agilent-0 All Entre CodeLine Description (Optional):	16436 Human miRNA Microarray 1.0 (CB 19116 Human miRNA Microarray 2.0 G44 z Gene IDs numan Whole Genome Bioarray	V1) 70B (CBI v1)	*	of proteinuria in which the primary ojects. Of which a subset of 11- thway analysis suggests common comparisons and answer key right within the application to choose
(Up to 5	00 characters)			
		The pre	ss "validate"	
				ly to Excel, PowerPoint and SVG.
Done			ernet 100% -	

# Concepts Analysis Upload

≁Upload My Concept			
Concept Name:	Podo-50-symbol		
Gene Set (Text File):		Browse	
	podocyte-50_gene symbol.txt		
Category:	HUGO Gene Symbol		
Null Set(s):	Affymetrix Human Genome HT U133 Plus 2.0 PM Array Agilent-016436 Human miRNA Microarray 1.0 (CBI v1) Agilent-019118 Human miRNA Microarray 2.0 G4470B (CBI v1) All Entrez Gene IDs CodeLink Human Whole Genome Bioarray		
Description (Optional):			
	(Up to 500 characters)		Upload Cancel
Concept [Podo-50-symbol] validated successfully 50 terms were recognized as distinct HUGO ger	ne symbols and will be uploaded.		1
			Then press "Upload"
Concont (Podo 50 c	(mbol) validated successfully		
Concept (Fouo-30-sy	indult valuated successfully		

# Concepts Analysis Upload

Concept Name: Podo-50-symbol	
Gene Set (Text File): podocyte-50_gene symbol.txt	
Category: HUGO Gene Symbol	
Null Set(s): All Entrez Gene IDs	
Description (Optional):	
Your custom concept [Podo-50-symbol] was successfully uploaded and can now be viewed in My Concepts. Select [Podo-50-symbol] as primary concept now.	Close
Concept (Podo-50-symbol) was successfully uploaded and can be viewed in My Concepts	
	-
Select (Podo-50-symbol) as primary concept now	

elect (Podo-50-symbol) as primary concept now

# **Concepts Summary View**

## Nephromine Concept Summary



Other (Non-Nephromine) Concept Summary

# **Concepts Analysis**



# **Concepts Analysis**

datasets	
----------	--

### concepts

ORDER BY: Dataset Name \$

ON: 🔶 🌲

\_\_\_\_ Compare | Clear All

- Hodgin FSGS (30)
- Group: Collapsing Focal Segmental Glomerulosclerosis vs. Focal Segmental Glomerulosclerosis
  - Group: Collapsing Focal Segmental Glomerulosclerosis vs. Minimal Change Disease and Normal Kidney
  - Group: Collapsing Focal Segmental Glomerulosclerosis vs. Normal Kidney
- Group: Focal Segmental Glomerulosclerosis vs. Minimal Change Disease and Normal Kidney
  - Group: Focal Segmental Glomerulosclerosis vs. Normal Kidney
  - Group: Minimal Change Disease vs. Normal Kidney

### visualize

Differential Analysis

GROUP BY: Group

SHOW: Only Samples in Analysis

#### 1 | 2 | 3 -

Comparison of Concept: "Podo-50-symbol - My Conc Under-expression in Group: Collapsing Focal Segmental Glomerul (log2 median-centered intensity) PowerPoint Publication-quality graphic (SVG) Excel - Analysis Comparison Excel - Analysis Gene List Excel - Dataset Detail

Rank	P-value	Fold Change	Gene		Reporter
99	0.001	-2.02	ACTN4		g3157975_3p_at
100	0.001	-3.74	SYNPO		g6005797_3p_at
142	0.002	-1.75	MAGI2		Hs.229355.0.A1_3p_at
156	0.002	-2.07	TJP1		g4507516_3p_at
171	0.002	-2.56	PODXL		g4885556_3p_at
194	0.002	-2.63	CLIC5		g8393146_3p_at
234	0.003	-2.78	NES		g13375818_3p_at
272	0.003	-2.19	SULF1		Hs.70823.0.\$3_3p_at
278	0.003	-2.18	NPHS1		207673_3p_at
359	0.004	-1.25	LRRC7		Hs2.157325.1.S1_3p_s_at
385	0.005	-2.67	TCF21		g4507394_3p_at
504	0.006	-2.46	NPHS2		g7657614_3p_at
580	0.008	-1.58	DAG1		g4758115_3p_a_at
590	0.008	-3.31	PLCE1		g7705940_3p_s_at
594	0.008	-1.57	FYN		g181171_3p_a_at
658	0.009	-1.97	WT1		g13386509_3p_a_at
696	0.009	-2.99	EZR		g340216_3p_a_at
707	0.009	-1.13	CD80		Hs2.838.3.51_3p_at
717	0.009	-2.39	MAFB		Hs.169487.0.51_3p_a_at
747	0.010	-2.40	CD 2AP		g11321633_3p_at
				1 2	

#### Legend

1. Normal Kidney (9)

2. Collapsing Focal Segmental Glomerulosclerosis (6)

## tranSMART

# The Translational Challenge: Data Integration & Analysis





# tranSMART Platform: Enabling Translational research



## tranSMART – A platform and community

- Open-source and opendata translational biomedical research community
- Biomedical Researchers, Developers, Service Providers
- Clinician Researchers

# tranSMART Platform: Academics and industry

2012 St

2009 Johnson and Johnson		2010 Thomson Reuters		2012 One Mind for Research		Jude, <u>Harvard</u> , Johns Hopkins Univ.
	2010 Sage Bionetw orks		2012 FDA		2012 Pfizer	

# tranSMART: controlled vocabulary



# Subset selection

← → C ↑ C ↑ C ↑ C ↑ C ↑ C ↑ C ↑ C ↑ C ↑ C										
Search Dataset Explorer Gene Signatur	e/Lists Cross-Database F	Exploration Admin								
Search Terms Navigate Terms Across Trials	🎭 Generate Summary Statistics   📄 Summary     🍘 Clear   🔚 Save									
0	Comparison Advanced Workflo	w Results/Analysis Grid View	Data Export Export Job	5						
	Subs	et 1 Exclude X	1	Subset 2	Exclude X					
Clinical Measurements (55) Clinical Measurements (55) Can further Can further Can further (43Specify with	\FSGS\ \eGFR v2l	ND Exclude X	\MCD\ \eGFR v2\	AND	Exclude X					
123 eGFR v2 (50) 123 eGFR v4 (42) AND or 123 eGFR v5 (43) 123 eGFR v6 (38) exclusion 123 eGFR v7 (35) 123 eGFR v8 (20)	A	ND Exclude X		AND	Exclude X					
	Subset 1			Subset 2	]					

## Summary statistics 1



Race

0%

100%

null

Total

0

11

Race

0%

100%

null

Total

0

5

ASIAN/

# Differentially expressed genes



#### Table of top Markers

													• • • • • • • • • • • • • • • • • • •						
Gene Symbol 🛛 🗢	Probe ID 🔶	Raw p-valute	Bonferro <b>‡</b> i	Holm	Hochber	SidakS#S	SidakS#	BH 🗢	BY \$	t \$	t (permutatior)	Raw P (permutation)	Adjusted P (permutation+)	Rank	S1 Meanŧ	S2 Mean 🖨	S1 SD 🖨	S2 SD \$	Fold Change
CENPE	A_16_P16795940	0.00000	0.00037	0.00037	0.00037	0.00037	0.00037	0.00018	0.00209	-5.789798	-5.789798	0.0004578755	0.2426740	1	0.38883375	-0.585704500	0.4883929	0.1823082	-0.66387358
LTF	16952883	0.00000	0.00037	0.00037	0.00037	0.00037	0.00037	0.00018	0.00209	-5.789798	-5.789798	0.0004578755	0.2426740	2	0.38883375	-0.585704500	0.4883929	0.1823082	-0.66387358
CORO1A	16817824	0.00001	0.40718	0.40716	0.40715	0.33447	0.33446	0.09503	1.00000	-4.469327	-4.469327	0.0011446886	0.8759158	3	0.63605159	-0.384358980	0.4674378	0.4016459	-1.65483734
ERC2	A_16_P16234993	0.00001	0.40718	0.40716	0.40715	0.33447	0.33446	0.09503	1.00000	-4.469327	-4.469327	0.0011446886	0.8759158	4	0.63605159	-0.384358980	0.4674378	0.4016459	-1.65483734
chr5:095910759-095910818	A_16_P17221956	0.00001	0.57016	0.57011	0.57010	0.43456	0.43454	0.09503	1.00000	-4.396770	-4.396770	0.0006868132	0.9063645	5	0.72159773	-0.020050546	0.4414995	0.2316556	-35.98893154
IGLV310	16932960	0.00001	0.57016	0.57011	0.57010	0.43456	0.43454	0.09503	1.00000	-4.396770	-4.396770	0.0006868132	0.9063645	6	0.72159773	-0.020050546	0.4414995	0.2316556	-35.98893154
MAP7	A_16_P17737416	0.00002	1.00000	1.00000	1.00000	0.68246	0.68242	0.14339	1.00000	4.242481	4.242481	0.0052655678	0.9553571	7	-0.05458949	0.881133820	0.5509338	0.3244491	-0.06195369
RN5S473	16866280	0.00002	1.00000	1.00000	1.00000	0.68246	0.68242	0.14339	1.00000	4.242481	4.242481	0.0052655678	0.9553571	8	-0.05458949	0.881133820	0.5509338	0.3244491	-0.06195369
AK092155	A 16 P17061075	0.00004	1.00000	1.00000	1.00000	0.90285	0.90282	0.23315	1.00000	4.080551	4.080551	0.0050366300	0.9809982	9	-0.06729611	0.450180580	0.3635915	0.1425495	-0.14948691

### Table of top Markers

### Enlarged:

Gene Symbol 🔶 🗘	Probe ID 🔶	Raw p-value	Bonferro#i
CENPE	A_16_P16795940	0.00000	0.00037
LTF	16952883	0.00000	0.00037
COR01A	16817824	0.00001	0.40718
ERC2	A_16_P16234993	0.00001	0.40718
chr5:095910759-095910818	A_16_P17221956	0.00001	0.57016
10000000000	1000000000	0.0000000	

P-values

Fold change

# Comparisons can be saved/emailed

← → C f L transmart-nephro.med	l.umich.edu:7070/transmart/datasetExplorer/index							
Search Dataset Explorer Gene Sign	nature/Lists Cross-Database Exploration Admin							
Search Terms Navigate Terms Across Trials	😵 🧐 Generate Summary Statistics   📄 Summary     🍘 Clear   🔚 Save							
0	Comparison Advanced Workflow Results/Analysis Grid View Data Export Export Jobs							
🔄 😋 Private Studies								
🖃 🔄 NeptunePOC2 (55)	Analysis: Heatmap							
🖃 😁 Biomarker Data (55)	Saved Comparison							
🖃 🔄 ST2 1 (55)	Cohorts:							
Kidney tub (55)	Subset 1: (\Private Studies\NeptunePOC2\Subjects\Medical History\Disease\dx\FSGS\ )							
G Clinical Measurements (55)	(\Private Studies\NentunePOC2\Clinical Measurements\Observations\eGER\eGER v2\ )							
122 everor (52)	Subset 2: (\Private Studies\NeptunePOC2\Subjects\Medical History\Disease\dx\MCD\) AND							
123 everyon (52)								
Observations (55)	(\Private Studies\NeptunePOC2\Clinical Measurements\Observations\eGFR\eGFR v2\ )							
	Variable Selection 2							
123 eGFR Slope (43)								
-123 eGFR v2 (50)	Heatmap Variable							
-123 eGFR v4 (42)								
123 eGFR v5 (43)	Select a High Dimensional Data node from the Data							
-123 eGFR v6 (38)	Set Explorer Tree and drag it into the box.							
-123 eGFR v7 (35)	X							
-123 eGFR v8 (20)	\kidney tub\							
-123 eGFR v9 (7)								
🖃 😋 Serum Creatinine (53)	High Dimensional Data							
-123 Screat v2 (53)								
123 Screat v4 (42)	Max rows to display : 50							
123 Screat v5 (43)								

# tranSMART – why do we care?

- Enables data exploration with low hurdles
- Integrates many different data types
- Has interfaces to real analysis tools
- Provides a consistent data set
- Can be run locally/ institutional etc
- Can possibly be "shared" across institutions
  - McMurry et al, PLOS one: Shrine: enabling nationally scalable Multisite disease studies
- Go to: <a href="http://transmartfoundation.org/">http://transmartfoundation.org/</a>

# Acknowledgements



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Matthias Kretzler Felix Eichinger Wenjun Ju Sebastian Martini Viji Nair Celine Berthier Laura Mariani Becky Steck Colleen Kincaid-Beal Rachel Dull Daniel R. Rhodes Rodney Keteyian Becky Steck Colleen Kincaid-Beal Rachel Dull

# Homework for fun

- Connectivity map
  - Use Diabetes vs. control (tubulointerstitium dataset)
  - Select top 1% overexpressed as primary concept
  - Compare to significantly overlapping concepts with Connectivity map
  - Can you find potential drug candidates? Are there any drugs that work for both glom. and tub?
  - What could be optimized? How will you plan further experiments to test your hypothesis?

